



Mini Review

Damage potential of *Tyrophagus putrescentiae* Schrank (Acari: Acaridae) in Oyster (*Pleurotus sajor caju*) Mushrooms

Komal Duhan, Rachna Gulati, Arvind Malik, Manoj

Abstract

Mite pests are cosmopolitan and reported from many parts of the world. *T. putrescentiae* was previously described as saprophagous species that feeds on different fungi including moulds (*Eutorium*, *Penicilium*) *Fusarium*, *Alternaria*, *Geotrichum*, *Mucor* and *Trichophyton*. Oyster Mushroom is a low investment, low risk, low labour intensive, and moderate profit enterprise. Mites have been found to damage mushrooms right from spawning to harvest of the crop. Moreover, mites themselves pollute the mushrooms by the multitude of faeces that they excrete all over, making the mushrooms uneatable as a food, causes allergies and other diseases. Considering the above facts, efforts have been made to assemble the available literature in this article.

Keywords faeces, harvest, Saprophagous, *T. putrescentiae*

Introduction

Mushrooms are valuable health foods that are low in calories, high in vegetable proteins, chitin, iron, zinc, copper, fibers, essential amino acids and vitamins [1]. Despite their nutritional and medicinal values, these are attacked by a number of diseases, pests, microbial infections and viruses that might infest the fungal substrate, the mycelium and the fruiting bodies growing on the substrate. Mushroom farms are generally infested by sciarid (*Lycoriella mali*), phorid (*Megaselia halterata*) and cecid (*Mycophila speyeri*, *Heteropeza pygmaea*) flies. The larvae of sciarid flies attack compost, spawn, mycelium, and pinhead formation. Cecid larvae feed on mushroom stems or gills and adult flies carry disease causing organisms to spread diseases. Three mycophagous nematode species, namely *Aphelenchoides composticola*, *Aphelenchus avenae* and *Ditylenchus myceliophagus* along with some saprophagous (*Rhabditis* sp.) and predatory (*Seinura* sp.) nematodes were found in compost samples of white button mushroom (*Agaricus* spp.) and oyster mushroom (*Pleurotus* spp.). The saprophagous nematodes (*Rhabditis* sp.) were recorded as the predominant species with 100 percent frequency of occurrence in oyster mushroom [2]. Several types of mite species had been found to be associated with mushrooms, out of which *Histiogaster* sp., *Histiostoma* sp., *Tarsonemus myceliophagus* *Tyrophagus lantneri*, *Uroobovella* sp., *Tarsonemus dimidatus*, and *Caloglyphus mycophagus* are of frequent occurrence [3]. Reports are available on the occurrence of mites including *T. putrescentiae* as pest in oyster mushrooms [4-5]

Host Preference of *Tyrophagus putrescentiae*

The mould mite, *Tyrophagus putrescentiae* (Schrank) has a worldwide distribution and is found in a variety of habitats including cultivated

Received: 2 March 2018

Accepted: 15 May 2018

Online: 21 May 2018

Authors:

Komal Duhan ✉, Rachna Gulati,
Arvind Malik, Manoj
Department of Zoology, Chaudhary Charan
Singh Haryana Agricultural University,
Hisar, India

✉ khushiduhan94@gmail.com

Emer Life Sci Res (2018) 4(1): 40-44

E-ISSN: 2395-6658

P-ISSN: 2395-664X

DOI: <http://dx.doi.org/10.7324/ELSR.2018.414044>

mushrooms, plant seeds, green houses, soil, nest of different animals, decaying organic material and foods containing high protein and fat content with moisture in the range of 12-15 percent [6-10]. Additionally, *T. putrescentiae* feeds on different fungi including moulds (*Eutorium*, *Penicilium*) *Fusarium*, *Alternaria*, *Geotrichum*, *Mucor* and *Trichophyton* [11-15]. It showed that collected pollen was an attractive food for *T. putrescentiae* [16]. It is very common in human created habitats such as dust in urban environments, medical and laboratory facilities, farms and food industry [11-12]. It prefers grain germ, pulses, nuts, sunflower, oil rape seeds, cheese, ham and dry dog food [17-20].

This species although prefer moist conditions and feeds on soil but damage stored produces due to their ability to tolerate low humidity and a wide range of temperatures [21-23] investigated allergic mites in dust and debris samples from 135 coal mines and most of them were *Tyrophagus putrescentiae*. *T. putrescentiae* is considered as an unpleasant pest damaging fungal cultures, but in the future, the use of this species in biological control of nematodes may be considered. All active developmental stages (larvae, nymphs and adults) of *T. putrescentiae* are nematophagous [24-25]. This mite avidly consumes nematodes including *Aphelenchus avenae* and *Meloidogyne javanica* as a food resource [16, 18, 22, 26-27].

***Tyrophagus putrescentiae* and other pest of mushrooms**

Use of old manure in compost, poor hygiene, sanitation in mushroom houses [28] and Dipterans flies as vectors for mites [22] are responsible for mite infestation in cultivated mushrooms. In India, seventy species of mites are associated with stored products including mushroom have been reported. The important pest species are *Acarus* sp., *Tyrophagus* sp., *Suidasia* sp., *Glycyphagus* sp., *Lardoglyphus* sp. and *Lepidoglyphus* sp. [12]. Among the different groups, Astigmata constitutes a dominant group in stored products ecosystem followed by Prostigmata and Mesostigmata [29]. Fifty four mite species had been found associated with mushrooms [30] out of which *Histiogaster* sp. [29, 31] *Histiostoma* sp. *Tarsonemus myceliophagus*, *T. lantneri*, *Caloglyphus mycophagus* *T. putrescentiae*, *Uroobovella* sp. were considered economically important pests. *T. putrescentiae* is one of the potential vectors of parasitic fungi such as *Aspergillus flavus*, *A. niger*, *Mucor racemosus* and *Nectria haematococca* which may cause economic damage in mushroom beds. *Oppia* sp. *Hypoaspis* sp. [32-34] *fimicolus* *Bakerdania* sp., *Luciaphorus* sp., *Parasitus* spp.[35-36], *Parasitus bituberosus* *Parasitus consanguineus* [37-38] were found to be associated with cultivated mushrooms.

Among the insect pests, sciarid flies Hump backed flies, cecid flies, springtails [37-38] and beetles [39-40] are predominant pests. Twenty-one nematode species representing two orders, *Aphelenchida* and *Tylenchida* have been reported to cause damage to mushroom cultivation in various parts of the world. Out of these 20 belong to four genera, *Aphelenchoides*, *Aphelenchus*, *Paraphelenchus* and *Seinura* under the order *Aphelenchida* and one species *Ditylenchus myceliophagus* belongs to the order *Tylenchida*. In India eight species of *Aphelenchoides* and *Ditylenchus myceliophagus* has been recorded from mushroom beds. Three *mycophagous* nematode species, i.e *Aphelenchoides composticola*, *A. avenae* and *Ditylenchus myceliophagus* along with some *saprophagous* (*Rhabditis* sp.) and predatory (*Seinura* sp.) nematodes were found in compost samples in villages of Sonipat district, Haryana, India. [12-13].

Abiotic stresses on mite development

Mites are reported from mushroom beds throughout the season. Jiang et al. [41] reported the presence of mites from September to March on temperate mushroom whereas they recorded mites in mushroom compost from April to June.

Temperature is an important factor affecting the development of mites. At favorable temperatures and 90 to 100 percent relative humidity, the *T. putrescentiae* female can lay an average of 437 eggs [42-43]. This revealed that temperature is an essential factor controlling the developmental time and life cycle of *Tyrophagus similis*. Nagesh and Reedy [34] showed that in wet condition, mold mites in the 20°C and 25°C temperature range remained alive for 31 days with no food. Under moist conditions (12 -18 % moisture) and warm summer temperature, a generation can be completed in 8 to 21 days. As the temperature falls, the length of the life cycle increases greatly. The mold mite breeds readily above 30 °C. The mold mite is less tolerant to low temperature and cannot develop below 10 °C. However, in an inactive state, this mite can

survive 0°C. The optimum temperature for the development and reproduction of *T. putrescentiae* and *Aleuroglyphus ovatus* is 25°C [11]. Low and high temperatures had negative effects on all immature stages and on the life cycle of both *T. putrescentiae* and *A. ovatus*. The duration of phorid development is temperature dependent and may vary between 15 days (24-27°C) to 50 days (16-21°C). The weight loss of the Shiitake mushroom caused by *T. putrescentiae* and *A. ovatus* increased with increasing temperature [9, 15].

Effect of infestation on biochemical composition of *Pluerotus sajor caju*

The mushroom mite, *Luciaphorus perniciosus* (Acari: Pygmeporidae) is the most important mite causing yield losses in mushroom production. According to Royse et al. [44] the most important factor in evaluating the nutritional value of mushrooms is their dry weight content, which generally lies between 5 and 15 percent of *P. sajor caju* fruit bodies. Variations in sugars content of mushroom might be due to variation in substrate and agroclimatic conditions. Maximum amount of carbohydrates (55.99/100gm) was found in *P. ostreatus* (white strain) cultivated on wheat straw, while the minimum amount of carbohydrates (35.26g/100g) was found in *P. ostreatus* (white strain) cultivated on cotton waste. Crude protein content of *P. sajor caju* fruit bodies varied between 27.4 and 34.8 percent. It is known that the protein content of mushrooms varies with the kind of substrate chosen, due to differential nature of the nutrient supply [25, 30, 32, 33].

Cabrera et al. and Gulati [45-47] reported that *T. myceliophagus*, *T. confuses* and *T. floreicolus* occur in thousands on mushroom body and the affected tissue becomes reddish, while base and stalk turn brown in colour. Accordingly *Pygmephorus* sp. feed on mushroom mycelium below the casing layer. Presence of the mites in the compost delayed the development of mushroom mycelium. Percent loss in total sugar, reducing sugar, non-reducing sugar, starch, protein content of compost was in the range of 10.47 to 24.60%, 22.89 to 68.67%, 0 to 0.92%, 16.33 to 33.33%, 0 to 10%, respectively due to *T. putrescentiae* infestation.

References

- [1] D. A. Ania, J. K. Oloke, S. G. Jonathan and O. J. Olawuyi (2012). Comparative assessment of mycelia biomass and exo polysaccharide production in wild type and mutant strains of *Scizophyllum commune* grown in Submerged liquid medium. *Nature Sci.*, **10**: 82-89.
- [2] A. U. Singh and K. Sharma (2016). Pests of Mushroom. *Adv. Crop. Sci. Tech.*, **4**: 213. doi:10.4172/2329-8863.1000213.
- [3] D. P. Tripathi (2005). *Mushroom Cultivation*, Oxford, IBH Publishing Co. Pvt. Ltd., New Delhi, p352.
- [4] C. D. Thapa and P. K. Seth (1982). Mushroom mites and their control. *Indian J. Mush.*, **8**: 45-52.
- [5] K. Okabe, K. Miyazaki and H. Yamamoto (2001). Population increase in mushroom pest mites on cultivated *Hypozygus marmoreus* and their vectoring of weed fungi between mushroom cultivation media. *JPN. J. APPL. ENTOMOL. Z.*, **45**: 75-81.
- [6] J. Arnau and L. Guerrero (1994). Physical methods of controlling mites in dry-cured ham. *Fleischwirtschaft (Germany)*, **74**: 1311-1313.
- [7] K. Solarz, P. Szilman and E. Szilman (1999). Allergenic mites associated with bird nests in Poland (Astigmata: Pyroglyphidae, Acaridae, Glycyphagidae). In *Ecology and Evolution of the Acari*, pp 651-656. Springer, Dordrecht, 1999.
- [8] L. Duek, G. Kaufman, E. Palevsky, I. Berdicevski (2001). Mites in fungal cultures. *Mycoses*, **44**: 390-394.
- [9] K. Kheradmand, K. Kamali, Y. Fathipour and E. M. Goltapeh (2007). Development, life table and thermal requirement of *Tyrophagus putrescentiae* (Astigmata: Acaridae) on mushrooms. *J. Stored. Prod. Res.*, **43**: 276-281.
- [10] P. Brazis, M. Serra, A. Selles, F. Dethioux, V. Biourge and A. Puigdemont (2008). Evaluation of storage mite contamination of commercial dry dog food. *Vet. Dermatol.*, **19**: 209-214.



- [11] F. Bahrami, K. Kamali and Y. Fathipour (2007). Life history and population growth parameters of *Tyrophagus putrescentiae* (Acari: Acaridae) on *Fusarium graminearum* in laboratory. Journal of Entomology Society of Iran, **26**: 7-18.
- [12] W. Chmielewski (1999). Acceptance of buckwheat grain as food by *Tyrophagus putrescentiae*. Fagopyrum, **16**: 95-99.
- [13] K. Solarz, L. Senczuk, H. Maniurka, E. Cichecka and M. Peszke (2007). Comparisons of the allergenic mite prevalence in dwellings and certain outdoor environments of the Upper Silesia (southwest Poland). Int. J. Hyg. Environ. Health, **210**: 715-724.
- [14] R. N. Sinha (1964). Effect of low temperature on the survival of some stored products mites. Acarologia, **6**: 336-341.
- [15] M. Singh and R. Gulati (2001). Minimization of losses due to mites in major pulses during storage. Final Technical Report. CSIR, New Delhi, 1-129.
- [16] P. L. Robertson (1961). A morphological study of variation in *Tyrophagus* (Acarina), with particular reference to populations infesting cheese. Bull. Entomol. Res., **52**: 501-529.
- [17] A. M. Hughes (1976). The mites of stored food and houses (No. 2nd edition). Technical Bulletin of the Ministry of Agriculture, Her Majesty's Stationery Office.
- [18] N. Garcia (2004). Efforts to control mites on Iberian ham by physical methods. Exp. Appl. Acarol., **32**: 41-50.
- [19] G. R. Anita, H. D. Kaushik and Arvind (2014). Efficacy of *Ocimum sanctum* and *Glycyrrhiza glabra* against stored mite, *Tyrophagus putrescentiae* Schrank in oat flakes. Biopesticides International, **10**: 41-49.
- [20] T. Erban, D. Rybanska, K. Harant, B. Hortova and J. Hubert (2016). Feces derived allergens of *Tyrophagus putrescentiae* reared on dried dog food and evidence of the strong nutritional interaction between the mite and *Bacillus cereus* producing protease bacillolysins and exo-chitinases. Front. Physiol., **7**: 53. doi: 10.3389/fphys.2016.00053.
- [21] K. K. Walia and S. Mathur (1995). Predatory behaviour of two nematophagous mites, *Tyrophagus putrescentiae* and *Hypoaspis calcuttaensis*, on root-knot nematode, *Meloidogyne javanica*. Nematol. Medit., **23**: 255-261.
- [22] D. E. Walter, R. A. Hudgens and D. W. Freckman (1986). Consumption of nematodes by fungivorous mites, *Tyrophagus* spp. (Acarina: Astigmata: Acaridae). Oecologia, **70**: 357-361.
- [23] S. Kumar, Y. Gautam and S. R. Sharma (2004). Mushroom mites and their management. Mushroom Res., **13**: 46-52.
- [24] A.D. Clift and R. B. Toffolon (1981). Biology, fungal host preferences and economic significance of two pygmephorid mites (Acarina: Pygmephoridae) in cultivated mushrooms, NSW, Australia. In Proceedings of the Eleventh International Scientific Congress on the Cultivation of Edible Fungi, Australia, 1981/edited by NG Nair, AD Clift. Sydney.
- [25] M. A. Abdel-Sater, S. K. Hemida, S. A. Eraky and M. M. Nasser (1995). Distribution of fungi on two mite species and their habitats in Egypt. Folia Microbiol., **40**: 304-313.
- [26] O. Ali, R. Dunne and P. Brennan (1997). Biological control of the sciarid fly, *Lycoriella solani* by the predatory mite, *Hypoaspis miles* (Acari: Laelapidae) in mushroom crops. Syst. Appl. Acarol., **2**: 71-80.
- [27] E. S. Binns (1973). Predatory mites neglected allies. Mushroom Journal, **12**: 540-545.
- [28] O. Ali, R. Dunne and R. Brennan (1999). Effectiveness of the predatory mite *Hypoaspis miles* (Acari: Mesostigmata: Hypoaspidae) in conjunction with pesticides for control of the mushroom fly *Lycoriella solani* (Diptera: Sciaridae). Exp. Appl. Acarol., **23**: 65-77.
- [29] E. Dmowska, J. Dmoch and K. Ilieva (1997). Early interaction of the bacterivorous nematode *Rhabditis cucumeris* and the edible fungus *Agaricus bisporus* in relation to time and quantity of nematode inoculation. International Journal of Mushroom Sci., **2**: 15-23.
- [30] Y. Chen (1967). The fungi of wheat straw compost: II. Biochemical and physiological studies. Trans. Br. Mycol. Soc., **50**: 667-677.



- [31] R. S. Gill, G. S. Sandu and M. S. Dhooria (1988). Studies on the pest and predatory status of mite, *Parasitus consanguineus* Oudemans and Voigts (Parasitidae: Mesostigmata) occurring on mushroom in Punjab. Indian J. Mush., **14**: 32-36.
- [32] S. Kasuga and H. Amano (2000). Influence of temperature on the life history parameters of *Tyrophagus similis* Volgin (Acari: Acaridae). Appl. Entomol. Zool., **35**: 237-244.
- [33] J. G. Rodriguez and L. D. Rodriguez (1987). Nutritional ecology of stored-product and house dust mites. Nutritional ecology of insects, mites, spiders and related invertebrates. Wiley, New York, 345-368.
- [34] P. P. Reddy, M. S. Rao, R. P. Tewari, M. Nagesh (1999). Evaluation of oil cakes for their efficacy against mushroom nematodes in chicken manure. Pest Manag. Hort. Ecosyst., **5**: 50-53.
- [35] J. Pumnuan, A. Insung and A. Chandrapatya (2008). Acaricidal effects of herb extracts on the mushroom mites, *Luciaphorus perniciosus* Rack and *Formicomotes heteromorphus* Magowski. Syst. Appl. Acarol., **13**: 33-38.
- [36] A. Jeyasankar, N. Raja and S. Ignacimuthu (2010). Antifeedant and growth inhibitory activities of *Syzygium lineare* Wall. (Myrtaceae) against *Spodoptera litura* Fab. (Lepidoptera: Noctuidae). Curr. Res. J. Biol. Sci., **2**: 173-177.
- [37] V. P. Sharma and K. Rajesh (2005). Use of botanicals to manage Sepadonium Yellow Mould and obtain higher yield in button mushroom. Indian J. Mycol. Pl. Path., **35**: 257-259.
- [38] I. Sanchez-Ramos and P. Castanera (2000). Acaricidal activity of natural monoterpenes on *Tyrophagus putrescentiae* (Schrank), a mite of stored food. J. Stored Prod. Res., **37**: 93-101.
- [39] I. Sanchez-Ramos and P. Castanera (2003). Laboratory evaluation of selective pesticides against the storage mite *Tyrophagus putrescentiae* (Acari: Acaridae). J. Med. Entomol., **40**: 475-481.
- [40] G. Gurusubramanian, A. Rahaman, M. Samrah, S. Roy and S. Bora (2008). Pesticide usage pattern in tea ecosystem, their retrospect and alternative measures. J. Environ. Biol., **29**: 813-826.
- [41] Y. Jiang, T. J. Smith and E. L. Ghisalberti (1997). The effect of volatile metabolites of lipid peroxidation on the aggregation of redlegged earth mites *Halotydeus destructor* (Acarina: Penthaleidae) on damaged cotyledons of subterranean clover. J. Chem. Ecol., **23**: 163-174.
- [42] G. Venkateshwarlu, M. V. Chandravada and R. P. Tewari (1999). Volatile flavor components of some edible mushrooms (Basidiomycetes). Flavour and Fragr J., **14**: 191-194.
- [43] B. Xia, D. Luo, Z. Zou and Z. Zhu (2009). Effect of temperature on the life cycle of *Aleuroglyphus ovatus* (Acari: Acaridae) at four constant temperatures. J. Stored Prod. Res., **45**: 190-194.
- [44] D. J. Royse, J. E. Sanchez, R. B. Beelman and J. Davidson (2008). Re-supplementing and recasing mushroom (*Agaricus bisporus*) compost for a second crop. World J. Microbiol. Biotechnol., **24**: 319-325.
- [45] R. A. Freire, G. J. de Moraes, E. S. Silva, A. C. Vaz and R. de Campos Castilho (2007). Biological control of *Bradysia matogrossensis* (Diptera: Sciaridae) in mushroom cultivation with predatory mites. Exp. Appl. Acarol., **42**: 87-93.
- [46] A. R. Cabrera, R. A. Cloyd and E. R. Zaborski (2005). Development and reproductive of *Stratiolaelaps scimitus* (Acari: Laelapidae) with fungus gnat larvae (Diptera: Sciaridae), potworms (Oligochaeta: Enchytraeidae) or *Sancassania aff sphaerogaster* (Acari: Acaridae) as the sole food source. Exp. Appl. Acarol., **36**: 71-81.
- [47] R. Gulati (2002). Bioefficacy of *Curcuma longa* L. oil against *Tyrophagus putrescentiae* Schrank and *Suidasia nesbitti* Hughes in wheat. Pest Management and Economic Zoology, **10**: 115-119.